

## IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re application of:

Ian MacLachlan, *et al.*

Application No.: 09/243,102

Filed: February 2, 1999

For: SYSTEMIC DELIVERY OF  
SERUM STABLE PLASMID LIPID  
PARTICLES FOR CANCER THERAPY

Examiner: J. Zara

Art Unit: 1635

Declaration of Mark Murray Under 37  
C.F.R. §1.132Assistant Commissioner for Patents  
Washington, D.C. 20231

Sir:

I, Mark Murray, being duly warned that willful false statements and the like are punishable by fine or imprisonment or both, under 18 U.S.C. § 1001, and may jeopardize the validity of the patent application or any patent issuing thereon, state and declare as follows:

1. All statements herein made of my own knowledge are true and statements made on information or belief are believed to be true.

2. I hold a Ph.D. (1978) from the University of Oregon Health Sciences University, and a M.S. (1973) from the University of San Francisco. I am presently the President and Chief Executive Officer for Protiva Biotherapeutics, Inc. (Burnaby, Canada).

My field of expertise is molecular oncogenesis and protein therapy. I have authored nineteen publications in the fields of molecular oncogenesis and protein therapy. A true copy of my *Curriculum Vitae* is attached hereto as Exhibit A.

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3. The present invention is directed to methods of treating tumors in mammals by delivery of serum stable nucleic acid-lipid particles with a nucleic acid portion that is fully encapsulated within the lipid portion. The delivery of serum stable nucleic acid-lipid particles is by injection at an injection site that is distal to the tumor in the mammal. The lipid portion of the nucleic acid-lipid particle comprises a cationic lipid, a neutral lipid, and a lipid that prevents aggregation during formulation.

4. I have read and am familiar with the contents of the subject patent application. I have also read the Office Action received from the United States Patent and Trademark Office dated October 3, 2002. It is my understanding that the Examiner is concerned that the claimed methods (1) are not enabled by the specification; and (2) are anticipated. Specifically, the Examiner states that the Applicants have demonstrated (1) numerous examples of successful *in vitro* and *in vivo* delivery of nucleic acids using the nucleic acid-lipid particles describe herein and (2) the ability to treat various tumor models *in vivo* using the claimed methods. However, the Examiner alleges that the specification is not enabling for (1) treatment of any neoplasm by distal administration of any nucleic acid fully encapsulated within any nucleic acid-lipid particle and (2) nucleic acid-lipid particles comprising any lipid formulation.

5. This declaration is provided to demonstrate that practice of the claimed methods is fully enabled by the specification and that the claimed methods are not anticipated. This declaration presents (1) additional experiments in which nucleic acids fully encapsulated in lipid formulations are expressed in cells upon transfection; (2) additional experiments in which nucleic acid-lipid particles inhibited tumor growth after administration to mice seeded with tumors; and (3) multiple lipid formulations for use in nucleic acid-lipid particles. The results unequivocally demonstrate that (1) nucleic acids encapsulated in the nucleic acid-lipid particles of the present invention are effectively expressed for treatment of neoplasia; (2) administration of the nucleic acid-lipid particles of the present invention is effective for treating tumors; and (3) multiple lipid

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formulations can be used for the lipid portion of the nucleic acid-lipid particles. One of skill in the art can therefore practice the claimed methods using information provided in the specification, together with methodology known to one of skill in the art at the time of the present invention, with at most, only routine experimentation.

6. The models used in carrying out the experiments described below are art-accepted models for (1) measuring transfection of cells with nucleic acids and (2) treating tumors. As demonstrated by the examples, (1) nucleic acids encapsulated in the lipid portion of the nucleic-acid lipid particles of the present invention are expressed in tumor cells and (2) the methods of the present invention are effective for treating tumors by distal administration of expressible genes fully encapsulated within the nucleic acid-lipid particles disclosed and claimed in the patent application.

7. As discussed with the Examiners during the interview of June 11, 2003, one advantage of the nucleic acid-lipid particles is tumor specific expression of nucleic acids encapsulated in the lipid portion of the nucleic acid-lipid particle. Additional experiments which are not described in the specification demonstrate that nucleic acids fully encapsulated in the nucleic acid-lipid particles of the present invention are expressed in tumor cells, but not in normal tissues. Two sets of experiments were conducted and are described below.

In the first set of experiments, mice were seeded with tumor cells. The mice were then intravenously injected with a nucleic acid-lipid particle containing a plasmid encoding luciferase. The mice were sacrificed and expression of luciferase was determined. Luciferase expression was observed in the tumor cells, but no significant luciferase expressed was observed in liver, spleen, lung, or kidney. A graphic illustration of the results is shown as Exhibit B.

In the second set of experiments, mice were seeded with tumor cells. The mice were then intravenously injected with a nucleic acid-lipid particle containing a plasmid encoding luciferase. The mice were sacrificed and expression of luciferase was

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determined. Luciferase expression was observed in the tumor cells, but no significant luciferase expression was observed in spleen, liver, adrenal glands, small intestine, thymus, lymph node, testes, kidney, heart, lung, large intestine, bone, or brain. A graphic illustration of the results is shown as Exhibit C.

These experiments demonstrate that delivery of any nucleic acid encapsulated in the nucleic acid-lipid particles disclosed and claimed in the patent application will lead to specific expression of the nucleic acid in tumor cells.

8. Additional experiments which are not described in the specification demonstrate that nucleic acids fully encapsulated in the nucleic acid-lipid particles of the present invention are actually expressed. Four sets of experiments were conducted and are described below.

In the first set of experiments, mice were seeded with neuroblastoma cells. The mice were then intravenously injected with a nucleic acid-lipid particle containing a plasmid encoding luciferase. The mice were sacrificed and expression of luciferase was determined. Luciferase expression was observed in the neuroblastoma cells.

In the second set of experiments, mice were seeded with melanoma cells. The mice were then intravenously injected with a nucleic acid-lipid particle containing a plasmid encoding luciferase. The mice were sacrificed and expression of luciferase was determined. Luciferase expression was observed in the melanoma cells.

The third set of experiments, mice were seeded with glioblastoma cells. The mice were then intravenously injected with a nucleic acid-lipid particle containing a plasmid encoding luciferase. The mice were sacrificed and expression of luciferase was determined. Luciferase expression was observed in the glioblastoma cells.

In the fourth set of experiments, mice were seeded with fibrosarcoma cells. The mice were then intravenously injected with a nucleic acid-lipid particle containing a plasmid encoding luciferase. The mice were sacrificed and expression of luciferase was determined. Luciferase expression was observed in the fibrosarcoma cells.

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These examples unequivocally establish that administration of nucleic acid-lipid particles as claimed in the present invention leads to expression of the nucleic acid fully encapsulated in the nucleic acid-lipid particles.

9. In addition to the extensive examples in the specification showing that growth of multiple tumor types is inhibited or reduced by administration of nucleic acid-lipid particles using the presently claimed methods, results from continuing experiments demonstrate that the growth of additional tumor types are inhibited by delivery of the nucleic acid-lipid particles using the presently claimed methods. In particular, growth of neuroblastoma, K1735 melanoma, and Harding Passey melanoma have all been shown to be inhibited.

10. Three additional sets of experiments, which again use art-accepted tumor models, demonstrate the efficacy of the nucleic acid-lipid particles administered using the presently claimed methods. These experiments, which were not described in the specification, are described below.

In the first set of experiments, mice were seeded subcutaneously with Neuro 2A neuroblastoma cells. On day 5 and every other day following for eight days, empty nucleic acid-lipid particles or nucleic acid-lipid particles containing a nucleic acid encoding HSV-TK were intravenously administered to the mice. Beginning on day 5 and every day following, mice were treated intraperitoneally with lipid formulated ganciclovir once daily. Mice treated with HSV-TK in nucleic acid-lipid particles and ganciclovir exhibited a marked reduction in tumor growth rate. A graphic illustration of the results is attached as Exhibit D.

In the second set of experiments, mice were seeded subcutaneously with Harding Passey melanoma cells. On day 17 and every other day following for eight days, empty nucleic acid-lipid particles or nucleic acid-lipid particles containing a nucleic acid encoding HSV-TK were intravenously administered to the mice. Beginning on day 19 and every day following, mice were treated intraperitoneally with ganciclovir once daily.

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Mice treated with HSV-TK in nucleic acid-lipid particles and ganciclovir exhibited a marked reduction in tumor growth rate. A graphic illustration of the results is attached as Exhibit E.

In the third set of experiments, mice were seeded subcutaneously with K1735 melanoma cells. On day 12 and every other day following for eight days, empty nucleic acid-lipid particles or nucleic acid-lipid particles containing a nucleic acid encoding HSV-TK were intravenously administered to the mice. Beginning on day 14 and every day following, mice were treated intraperitoneally with ganciclovir once daily. Mice treated with HSV-TK in nucleic acid-lipid particles and ganciclovir exhibited a marked reduction in tumor growth rate. A graphic illustration of the results is attached as Exhibit F.

These experiments demonstrate that nucleic acid-lipid particles delivered according to the presently claimed methods are effective for inhibiting growth of additional tumor types when administered at a site distal to the tumor. In particular, these experiments demonstrate that, administration of fully encapsulated nucleic acids encoding are effective in inhibiting tumor growth.

11. As explained above, the lipid portion of the nucleic acid-lipid particles of the presently claimed invention comprises a cationic lipid, a neutral lipid and a lipid that prevents aggregation during formulation (*i.e.*, a fusion regulating lipid). Multiple cationic lipids, neutral lipids, and fusion regulating lipids can be used in the nucleic acid-lipid particles in which the nucleic acid is fully encapsulated in the lipid portion of the particle as disclosed and claimed in the present invention. The ratio of cationic lipids to neutral lipids to fusion regulating lipids can also be varied in the nucleic acid-lipid particles of the claimed invention. Suitable cationic lipids include, but are not limited to DODAC, DODMA and its derivatives, DOTAP, DOPC, and DC-Chol. Suitable neutral lipids include but are not limited to DOPE, DSPC, and cholesterol. Suitable lipids that prevent aggregation include, but are not limited to PEG-lipids such as PEG Ceramides, PEG-diacylglycerols, PEG-DSA derivatives as well as PEO lipid

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derivatives. Thus, multiple lipid formulations can be used in the nucleic acid-lipid particles of the presently claimed invention. For example, at least twenty five representative lipid formulations (including fourteen different options for cationic lipid components, seven different options for neutral lipid components, and eleven options for lipids that prevent aggregation during formulation) that have actually been used to make lipid portions of nucleic acid lipid particles used in the presently claimed invention. A chart summarizing these formulations is attached as Exhibit G.

12. It is my understanding that in the Office Action mailed October 3, 2002, the Examiner cited two references that allegedly anticipate the claimed methods: Kirn *et al.*, (U.S. Patent No. 6,133,243) and Hung *et al.* (U.S. Patent No. 6,197,754).

Kirn *et al.* disclose nucleic acid-lipid complexes. More particularly, Kirn *et al.* describe preparation of nucleic acid-lipid complexes by first preparing cationic liposomes. Only after the liposomes are fully formed are they mixed with nucleic acids to form nucleic acid-lipid complexes (*see, e.g.*, col. 11, line 65 to col. 12, line 61). Thus, in contrast to the nucleic-acid lipid particles used in the methods of the present invention, the nucleic acid-lipid complexes of Kirn *et al.* do not comprise a nucleic acid fully encapsulated in the lipid portion of a nucleic-acid lipid particle.

Hung *et al.* disclose nucleic acid-lipid complexes. More particularly, Hung *et al.*, describe preparation of nucleic acid-lipid complexes by first preparing cationic liposomes. Only after the liposomes are fully formed are they mixed with nucleic acids to form nucleic acid-lipid complexes (*see, e.g.*, col. 38, line 58 to col. 39, line 30). Thus, in contrast to the nucleic-acid lipid particles used in the methods of the present invention, the nucleic acid-lipid complexes of Hung *et al.* do not comprise a nucleic acid fully encapsulated in the lipid portion of a nucleic-acid lipid particle.

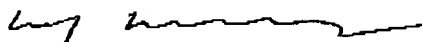
Thus, neither of the cited references anticipate the claimed methods of treating tumors in mammals by delivering serum stable nucleic acid-lipid particles comprising a nucleic acid portion that is fully encapsulated within the lipid portion.

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13. In view of the foregoing, it is my scientific opinion that one of skill in the art would be able to practice the claimed invention with, at most, routine experimentation using the guidance in the specification and what is known to those of skill in the art. The specification, therefore, fully enables the methods of the invention.

In addition, it is my scientific opinion that neither Kirn *et al.* nor Hung *et al.* describe methods of treating tumors in mammals by delivery of serum stable nucleic acid-lipid particles with a nucleic acid portion that is fully encapsulated within the lipid portion. Thus, neither of the cited references anticipate the claimed invention.



Dated: June 29, 2003 By: \_\_\_\_\_  
Mark J. Murray, Ph.D.

SF 1472666 v1



## Exhibit A

**Mark J. Murray, Ph.D.**1127 41<sup>st</sup> Ave. E.  
Seattle, WA. 98112**PROFESSIONAL EXPERIENCE****PROTIVA BIOTHERAPEUTICS INC.**  
*President and Chief Executive Officer***June 2000 - Present****STEM CELL PHARMACEUTICALS**  
*Vice President, Business Development***Jan - June 2000****XCYTE THERAPIES, Seattle, WA**  
*Vice President, Business Development***1997- Dec 1999****ZYMOGENETICS INC., Seattle, WA.**  
*Vice President, Strategic Business Development*  
*Senior Director, New Business Development*  
*Director, New Business Development*  
*R&D Manager, Wound Repair Program*  
*Project Leader*  
*Senior Scientist***1982 - 1997**  
**1995 - 1997**  
**1993 - 1995**  
**1989 - 1992**  
**1987 - 1989**  
**1983 - 1987**  
**1982 - 1983****GRADUATE ACADEMIC EXPERIENCE****Postdoctoral Fellow**  
Center for Cancer Research, Massachusetts Institute of Technology  
Dr. Robert A. Weinberg, Supervisor**1979 - 1982****Postdoctoral Fellow**  
University of Oregon Health Sciences University  
Dr. David Kabat, Supervisor**1978 - 1979****Ph.D. Candidate**  
University of Oregon Health Sciences University  
Dr. David Kabat, Supervisor**1975 - 1978****Masters Degree Candidate**  
Department of Biology, University of San Francisco  
Dr. C. Peter Flessel, Supervisor**1971 - 1973**

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Exhibit A  
Page 2 of 6**SCIENTIFIC PUBLICATIONS**

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Exhibit A  
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Ogawa, Y., S.J. Sawamura, B.M. Pratt, J.M. McPherson, M.J. Murray and G.A. Ksander. Transforming growth factor- $\beta$  induces accumulation of hyaluronate *in vivo*. American Society of Cell Biology, San Francisco, 1989. (invited lecture)

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#### **PENDING U.S. PATENT APPLICATIONS**

Murray et al., Biologically Active A-chain Homodimers, No. 08/412,551, Allowed.

Four pending U.S. Patent Applications related to PDGF

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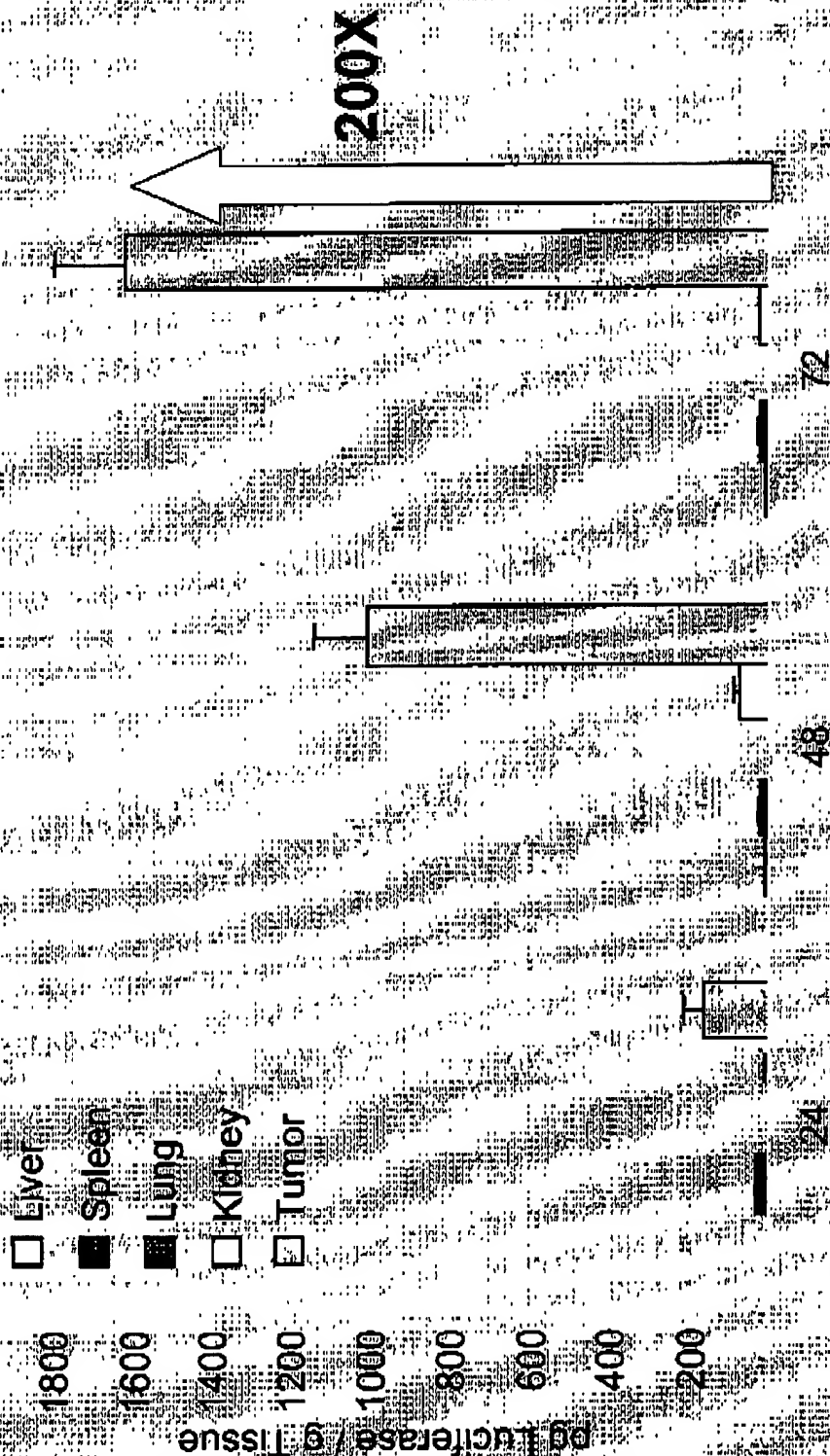
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Exhibit C

30

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0.5

ng Luciferase / g Tissue



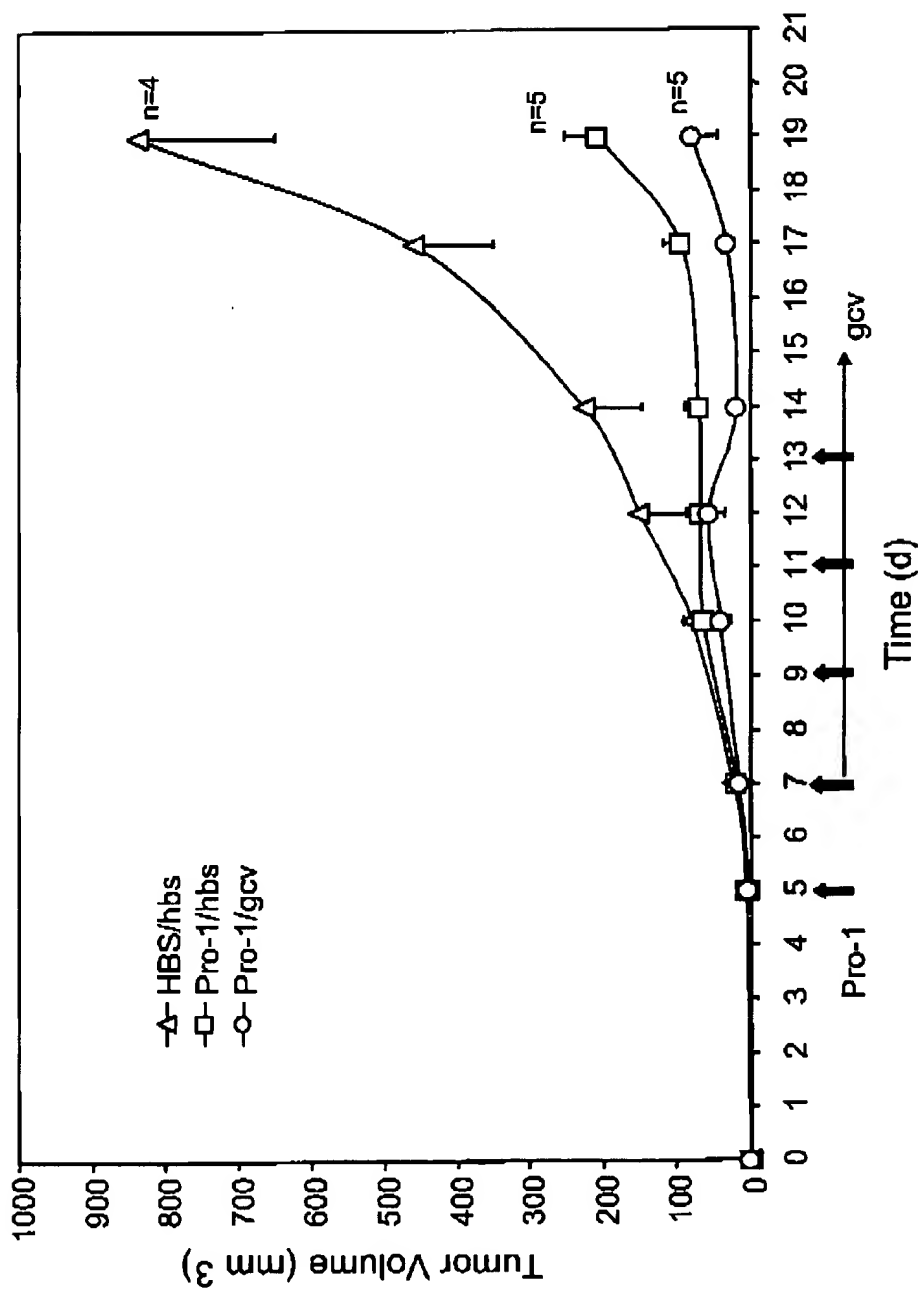
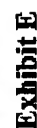


Exhibit D



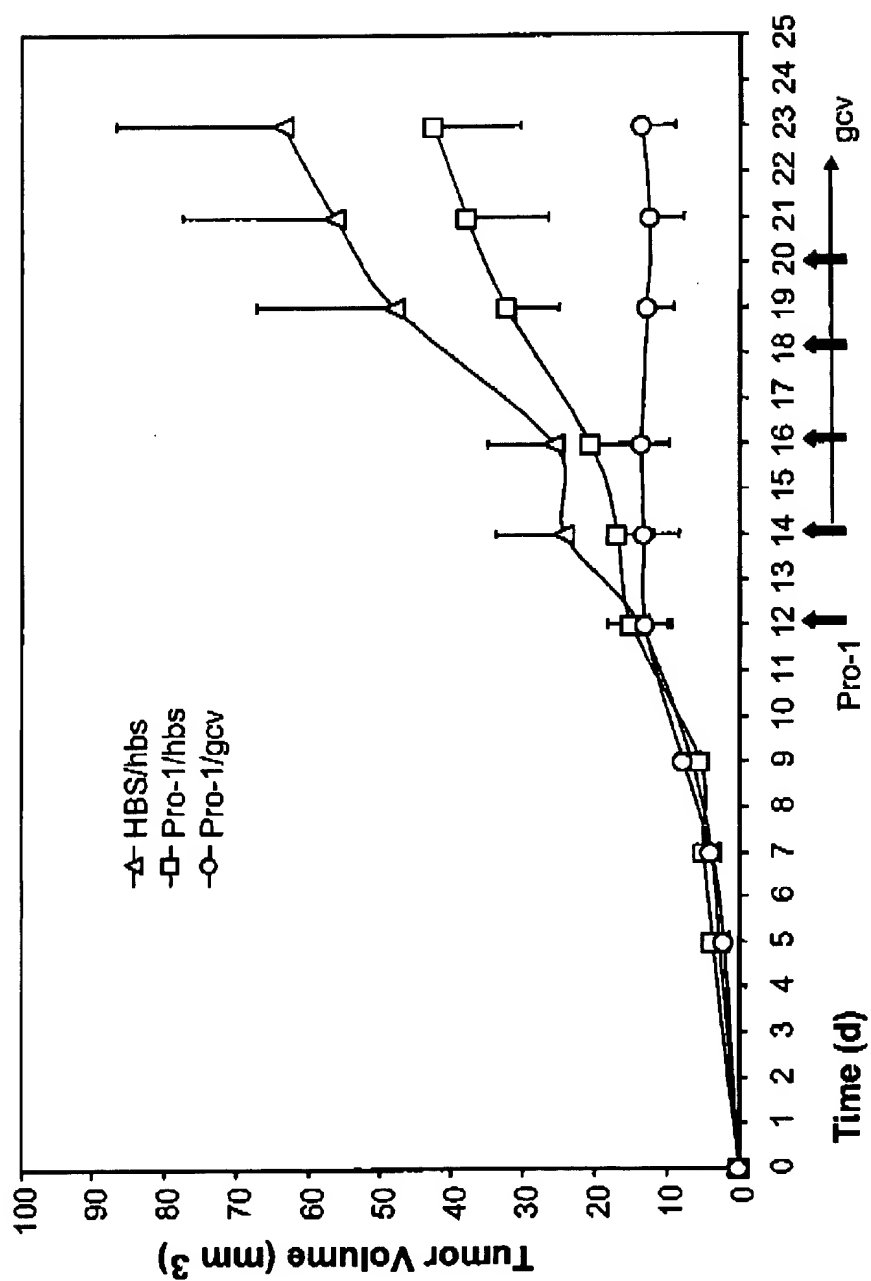


Exhibit F

## Exhibit G

#	Cationic Lipid(s) (mol%)	Neutral Lipid(s) (mol%)	Fusion Regulator(s) (mol%)
1	DODAC (7.5)	DOPE (82.5)	PEG-CeramideC20 (10)
2	DODAC (7.5)	DOPE (82.5)	PEG-DSG (10)
3	DODAC (7.5)	DOPE (82.5)	PEG-DPG (10)
4	DODAC (7.5)	DOPE (82.5)	PEG-DMG (10)
5	DODAC (12)	DOPE (78)	PEG-CeramideC20 (10)
6	ethyl-DOPC (8)	DOPE (82)	PEG-CeramideC20 (10)
7	DOTAP (8)	DOPE (82)	PEG-CeramideC20 (10)
8	DOTAP (12)	DOPE (78)	PEG-CeramideC20 (10)
9	DOTAP (16)	DOPE (74)	PEG-CeramideC20 (10)
10	DC-Chol (8)	DOPE (82)	PEG-CeramideC20 (10)
11	DC-Chol (12)	DOPE (78)	PEG-CeramideC20 (10)
12	DC-Chol (16)	DOPE (74)	PEG-CeramideC20 (10)
13	DC-Chol (20)	DOPE (70)	PEG-CeramideC20 (10)
14	DODMA-AN (8)	DOPE (82)	PEG-CeramideC20 (10)
15	DODMA-AN (12)	DOPE (78)	PEG-CeramideC20 (10)
16	DODMA (25)	DSPC (20) Cholesterol (45)	PEG-DSG (10)
17	DODMA (15)	DSPC (20) Cholesterol (55)	PEG-DSG (10)
18	DODMA (15)	DSPC (20) Cholesterol (55)	PEG-CeramideC20 (10)
19	DODMA (15)	DSPC (20) Cholesterol (55)	PEG-A-DMA (10)
20	DODMA (15)	DSPC (20) Cholesterol (55)	PEG-A-DPA (10)
21	DODMA (15)	DSPC (20) Cholesterol (55)	PEG-A-DSA (10)
22	DODMA (15)	DSPC (20) Cholesterol (55)	PEG-USPE (10)
23	DODMA (15)	DSPC (20) Cholesterol (55)	PEG-amide-DSA (10)
24	DODMA (15)	DSPC (20) Cholesterol (55)	PEG-carbamate-DSA (10)
25	DODMA (15)	DSPC (20) Cholesterol (55)	PEG-succinimide-DSA (10)